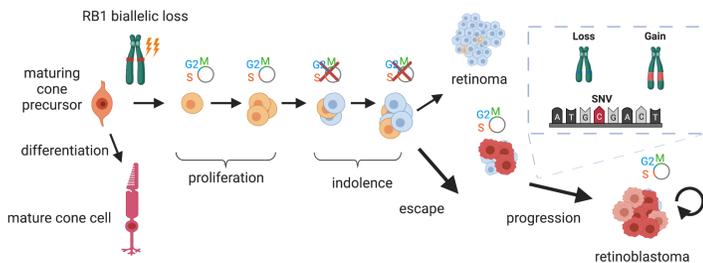


Background

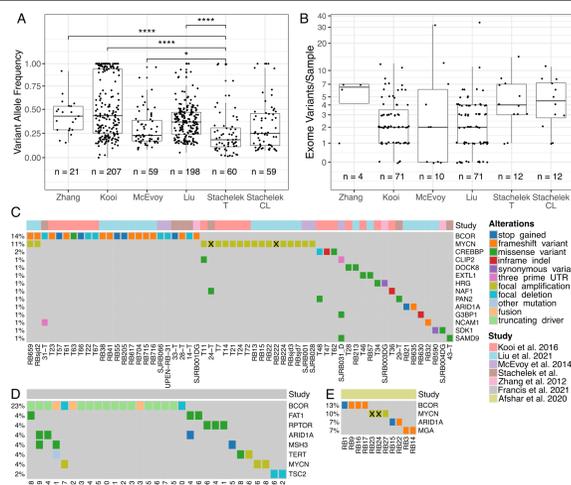
- Retinoblastomas initiate due to biallelic RB1 loss or MYCN amplification and progress through secondary mutations.
- These mutations include non-recurrent nucleotide variants whose significance has been unclear.
- Whole genome or exome sequencing (WGS/WES) of 156 treatment-naive retinoblastomas has revealed no significantly recurrently altered genes with exome tier-1 mutations



Approach

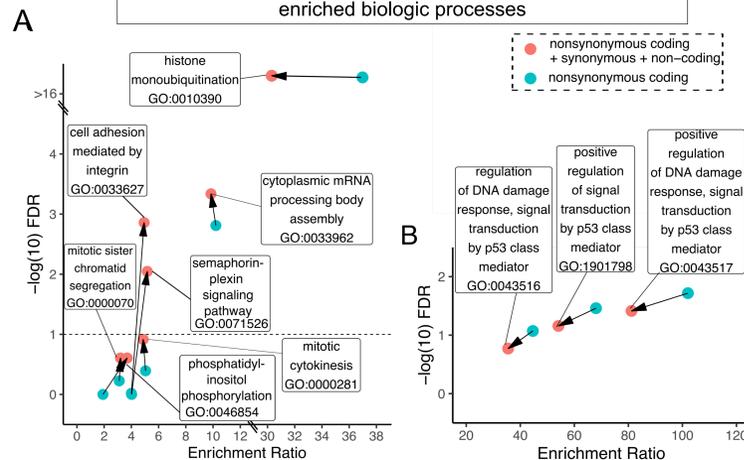
- Call somatic variants using GATK MuTect2 and Strelka2 in retinoblastoma tumors and cell lines relative to matched normal DNAs
- Identify recurrently altered progression-related processes by integrating whole exome sequencing of 12 retinoblastomas and matched cell lines with 156 retinoblastomas from prior studies.
- Determine contributions of noncoding and synonymous variants in overrepresented processes
- Identify transcriptional effects of 3' UTR sequences of *PCGF3* and *CDC14B* by luciferase reporter assay.
- Test post-transcriptional effects of synonymous mutation in *DYNC1H1* using CRISPR base editing and immunoblotting

Variant allele frequencies, variant numbers and mutated genes in retinoblastoma WES/WGS studies



Somatic variant frequencies and repeatedly mutated genes beyond RB1 in 168 fully reported retinoblastoma tumors. A. Variant allele frequencies (VAF) of all exomic non-RB1 single nucleotide and indel variants in this and four prior studies. B. Number of variants detected per sample in each study displayed with pseudo log scale. C-E. Repeatedly mutated genes beyond RB1 identified in retinoblastoma whole genome or whole exome sequencing (C) or in targeted sequencing (D, E). Tumors with focal MYCN amplification and no annotated RB1 lesion are indicated with x.

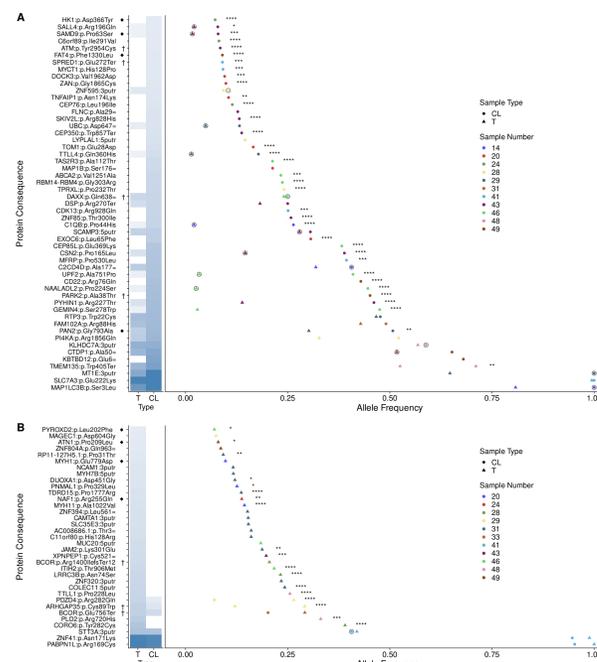
Non-coding and synonymous variants enhance detection of enriched biologic processes



geneSet	Ontology Description	FDR	Variant Genes in Retinoblastoma Tumors
GO:0010390	Histone monoubiquitination	< 1E-16	<i>BCOR</i> (18); <i>DDI1</i> ; <i>RNF20</i> ; <i>PCGF3</i>
GO:0016569	Covalent chromatin modification	2.25E-08	<i>BCOR</i> (18); <i>CREBBP</i> (2); <i>BRCA2</i> ; <i>BRMS1</i> ; <i>CHD1</i> ; <i>DDB1</i> ; <i>EHMT1</i> ; <i>EYA1</i> ; <i>HDAC10</i> ; <i>HIST1H1E</i> ; <i>KAT5A</i> ; <i>KDM8</i> ; <i>NSD1</i> ; <i>PADI4</i> ; <i>PRKCA</i> ; <i>RNF20</i> ; <i>SUPT6H</i> ; <i>TAF1</i> ; <i>TAF1L</i> ; <i>TAF9</i> ; <i>PCGF3</i> ; <i>HDAC9</i>
GO:0033962	Cytoplasmic mRNA processing (P) body assembly	4.60E-04	<i>PAN2</i> (2); <i>CNOT1</i> ; <i>CNOT2</i> ; <i>DYNC1H1</i>
GO:0022618	Ribonucleoprotein complex assembly	5.36E-04	<i>G3BP1</i> (2); <i>NAF1</i> (2); <i>PAN2</i> (2); <i>CELF3</i> ; <i>CNOT1</i> ; <i>CNOT2</i> ; <i>EIF3E</i> ; <i>EIF4B</i> ; <i>GEMIN1</i> ; <i>NLEF5</i> ; <i>TAF9</i> ; <i>DYNC1H1</i> ; <i>GEMIN8</i> ; <i>RPF2</i> ; <i>USP4</i>
GO:0033627	Cell adhesion mediated by integrin	1.38E-03	<i>HRO</i> ; <i>HRO3</i> ; <i>CRK</i> ; <i>ITGB4</i> ; <i>ITGB8</i> ; <i>ITGBL1</i> ; <i>ACER2</i> ; <i>ITGB2</i>
GO:0071526	semaphorin-plexin signaling pathway	9.00E-03	<i>NCAM1</i> (2); <i>SEMA3G</i> ; <i>SEMA4D</i> ; <i>NET</i>
GO:0000281	Mitotic cytokinesis	0.121	<i>BRCA2</i> ; <i>ANK3</i> ; <i>INCENP</i> ; <i>NUP62</i> ; <i>SEPT10</i> ; <i>SPTBN1</i> ; <i>STAMPB</i> ; <i>LUNC119</i> ; <i>MYH10</i> ; <i>ZFYVE26</i>
GO:0046854	phosphatidylinositol phosphorylation	0.248	<i>CISH</i> ; <i>FGF18</i> ; <i>PI4KA</i> ; <i>PIKFYVE</i> ; <i>ICOS</i> ; <i>MET</i> ; <i>PIK3CD</i> ; <i>PIK3R6</i> ; <i>PIPSK1C</i> ; <i>SEC</i>
GO:0000070	Mitotic sister chromatid segregation	0.248	<i>BOD1</i> ; <i>BUB1B</i> ; <i>DYNC1L1</i> ; <i>HIRA</i> ; <i>INCENP</i> ; <i>KIF14</i> ; <i>NUP62</i> ; <i>PIBF1</i> ; <i>PRC1</i> ; <i>CDC14B</i> (2); <i>RAB11A</i> ; <i>TTN</i>

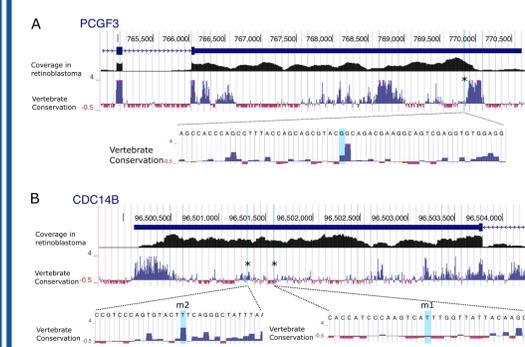
Biological process gene ontology terms over-represented in exome variants in retinoblastoma whole exome or whole genome sequencing. performed separately for non-synonymous variants affecting protein amino acid sequence (blue) and for all exomic variants including synonymous coding mutations and non-coding 5' or 3' UTR mutations (red)

Enrichment and depletion of somatic variants during retinoblastoma cell line establishment



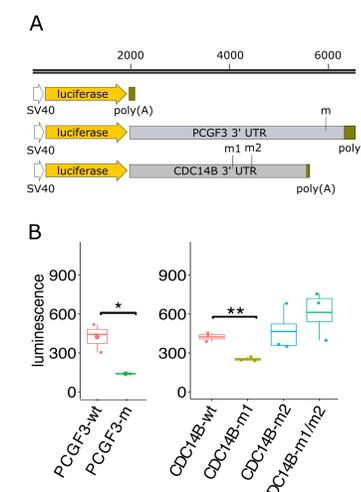
Curated somatic variants in CHLA-VC-RB tumor and cell lines. Variants in tumors (triangles) and cell lines (circles) are displayed in ascending VAF order for samples with increased variant frequency in tumor-derived cell lines. Circled icons indicate variants initially detected in a cell line and subsequently detected exclusively in the matched tumor. ♦, alleles confirmed by targeted sequencing. †, genes listed in targeted sequencing panels (UCSF500 or MSK-IMPACT). Asterisks, significantly altered allele frequencies in tumor-cell line pairs (*, p<0.05; **, p<0.01; ***, p<0.001; ****, p<0.0001; Fisher exact test).

Noncoding variants of interest in 3' UTR

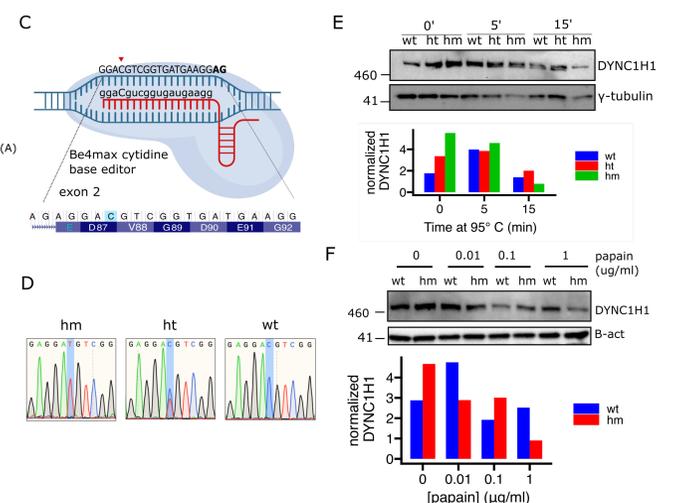


Expression and conservation of PCGF3 and CDC14B variant sequences. UCSC Browser views of PCGF3 (A) and CDC14B (B) 3' UTRs, displaying RNAseq expression in a representative retinoblastoma tumor (GSE125903) and vertebrate conservation at low and high resolution. Note genomic CDC14B sequences represent the cDNA minus strand. Asterisks and blue shading indicate variant positions

Noncoding variants in PCGF3 and CDC14B impact transcription



Synonymous variant in DYNC1H1 impacts post-transcriptional stability



Effects of non-coding and synonymous mutations in PCGF3, CDC14B and DYNC1H1. A. pGL3-SV40 luciferase reporter plasmids showing PCGF3 and CDC14B 3' UTRs and position of mutations (m, m1, m2). B. Luciferase assays in retinoblastoma cells transfected with the indicated reporter constructs. C. Base-editing strategy for DYNC1H1 c.261C>T using BE4max cytosine base editor. D. Sanger sequence traces of edited homozygous (hm), heterozygous (ht) and wild-type (wt) RB31 clones. E. DYNC1H1 immunoblot of whole cell lysates of homozygous and wild-type CHLA-VC-RB31 clones digested with papain at increasing concentrations. b-actin is a loading control. Graphs indicate DYNC1H1/b-actin ratios for each condition.

Conclusions

- Identified genomic abnormalities and altered cellular processes that contribute to retinoblastoma progression.
- Over-represented ontologies were more significantly over-represented when synonymous coding and non-coding UTR mutations were included
- Our analyses reveal that retinoblastoma progression is associated with accumulation of subclonal gene variants that affect cell signaling pathways different from those queried in current cancer gene sequencing panels.
- Most of the affected biological process ontologies were more significantly over-represented when including non-coding UTR variants that alter the levels and thus the stoichiometry of multi-protein complex components
- Noncoding and synonymous variants of unknown significance are demonstrated to impact transcription and post-transcriptional processes